

Comprehensive HRMS chemical characterization of pomegranate-based antioxidant drinks via a newly developed suspect and target screening workflow

Anthi Panara, Evangelos Gikas, Ilias Tzavellas and Nikolaos S. Thomaidis*

*Laboratory of Analytical Chemistry, Department of Chemistry, National and Kapodistrian University of Athens, 15771 Athens, Zografou GR-15771, Greece; panaranthi@chem.uoa.gr (A.P.); vgikas@chem.uoa.gr (E.G.); tzavell@chem.uoa.gr (I.T)

* Correspondence: ntho@chem.uoa.gr (N.S.T)

Abstract: Antioxidants play significant role in human health, protecting against a variety of diseases. Therefore, the development of products with antioxidant activity is becoming increasingly prominent in human lifestyle. New antioxidant drinks containing different percentages of pomegranate, black berries, red grapes, and aronia have been designed, developed, and manufactured by a local industry. The comprehensive characterization of the drinks' constituents has been deemed necessary to evaluate their bioactivity. Thus, LC-qTOFMS has been selected, due to its sensitivity and structure identification capability. The data dependent and independent acquisition modes have been utilized. The data have been treated according to a novel, newly designed workflow based on MS-DIAL and mzmine for suspect, as well as target screening. The classical MS-DIAL workflow has been modified to perform suspect and target screening in automatic way. Furthermore, a novel methodology based on compiled bioactivity driven suspect list was developed and expanded with combinatorial enumeration to include metabolism products of the highlighted metabolites. Compounds belonging to ontologies with possible antioxidant capacity have been identified, such as flavonoids, amino acids, and fatty acids, that could be beneficial to humans' health, revealing the importance of the produced drinks as well as the efficacy of the new in-house developed workflow.

Keywords:

Antioxidant drinks, novel workflows, HRMS, pomegranate, suspect screening methodology

Citation: Panara, A.; Gikas, E.; Tzavellas I. Thomaidis, N.S.

Comprehensive HRMS chemical characterization of antioxidant drinks via a newly developed suspect and target screening workflow

Molecules **2023**, *x*, x. <https://doi.org/xxxx>

Academic Editor: xxx

Received: date:

Accepted: date:

Published: date:

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2023 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Pomegranate (*Punica granatum L.*), classified as a berry, is a member of Pinaceae family and has been cultivated in the Mediterranean region (Turkey, Egypt, Tunisia, Spain), as well as in India and Iran [1]. Pomegranate has been highly valued, due to its nutritional and medicinal properties, as well as its biological and free radical scavenging activity, which are attributed to the antioxidant phytochemicals derived from various parts of the plant (peel, seed, leaf, and flower) [2-4]. Pomegranate juice is a rich source of polyphenols, fructose, carbohydrates, glucose and organic acids (i.e., ascorbic acid, citric acid, fumaric acid, and malic acid), while it contains several amino acids, including proline, methionine, and valine. Additionally, the presence of tannins and flavonoids, as the main type of polyphenols, indicates the pomegranate's pharmacological potential, due to their antioxidant activity [1]. Ellagic acid, a metabolized form of ellagitannin is a powerful antioxidant, and it has an extensive applicability in plastic surgery, preserving the viability of skin flaps.

Furthermore, anthocyanins (water-soluble pigments), flavan-3-ols, and flavanols are some of the flavonoids found in pomegranate related to plausible health benefits. Catechins, which can be found in both juice and the peel of pomegranate, are vital to the biosynthesis of anthocyanins and have antioxidant and anti-inflammatory properties. It should be noted that all the flavonoids appeared in pomegranate have antioxidant capacity and contribute to the indirect suppression of inflammatory indicators, such as tumor necrosis factor-alpha (TNF α) [1]. It has been demonstrated that pomegranate fruits can be utilized for the treatment of human prostate cancer inhibiting cell development and inducing apoptosis.

Aronia melanocarpa, commonly referred as "aronia," is a *Rosaceae* family plant that is native to eastern North America and has lately also been grown in Europe [5]. Aronia extracts demonstrated anti-oxidant, anti-diabetic, and anti-inflammatory activity [6]. *Morus nigra*, which belongs to the *Moraceae* family, is also known as black mulberry [7], and is cultivated in India and China, as well in the Mediterranean region in Greece and Turkey [8]. A number of biological activities, including antidiabetic, antioxidant, anti-inflammatory, and anti-hyperlipidemic, have recently been linked to mulberry fruit [9]. *Vitis vinifera* L. (red grapes), a member of *Vitaceae* family [10], has traditionally been located from South Caucasus toward Mediterranean basin [11], whereas more recently new countries (United States, Australia, China) have been involved in the cultivation of the species [12]. Grapes have known to possess a wide array of biological activities such as antioxidant, antimicrobial, anti-inflammatory, and anti-cancer properties [13].

Pomegranate's beneficial effect on human health resulted in an increasing growth in market share. Therefore, the global pomegranate market is expected to worth 248.4 million USD in 2022 and 338.6 million USD by 2028, with a CAGR of 5.3% during the review period [14]. In the term of this study, pomegranate-based juices were created with the addition of secondary ingredients; black berries, red grapes, and aronia. The selection of these ingredients, that could provide added value to the final drink, was based on their market availability, contribution to the antioxidant capacity of the juice, and effect on the flavor and inner flavor of the juice. Therefore, there is an increasing trend to introduce in the market these pomegranate-based drinks.

Regarding the evaluation of the drinks' value and the determination of its antioxidant capacity, two trends appear in the literature. The first approach is related to the application of conventional techniques (i.e., DPPH, TEAC assay, Folin-Ciocalteu method) and the second one is linked to the determination of the antioxidant content of juice in terms of molecular species. The first approach is prone to various limitations, due to the nature of the matrix with the most common being the interference from the background. To begin with, the DPPH (2,2-diphenyl-1-picrylhydrazyl) assay is a spectrometric method for determining antioxidants in solid or liquid matrices [15] via measuring substances' ability to act as free radical scavengers or hydrogen donors. This method can be employed specifically for the estimation of the overall antioxidant capacity and the free radical scavenging activity of fruit and vegetable juices [16]. Furthermore, Trolox Equivalent Antioxidant Capacity Assay (TEAC) measures spectrophotometrically the reduction of the radical cation ABTS⁺ (2,2-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid) by antioxidant compounds [17], and it is commonly used in scientific research for analyzing foods' and beverages' antioxidant capacity [18]. Another frequently employed method for the determination of the antioxidant capacity is the spectrophotometric method using the Folin-Ciocalteu reagent [19].

An alternative approach is the use of chromatography-based techniques for the determination of individual analytes that are connected to antioxidant activity. This approach ascertains the selectivity and credibility of the obtained results. High-pressure liquid chromatography with a photodiode array detector (HPLC-PDA) was used for the determination of organic and phenolic acids (11), and anthocyanins in juices (11, 12). In the direction to further enhance the sensitivity and specificity of the acquired results, the recent trend is towards mass spectrometry-based methodologies, i.e. performing LC-

MS/MS methods for the determination of organic acids [20], and HPLC/PDA/MS² for the quantification of punicalagin, ellagic acids and anthocyanidins [21]. A more comprehensive approach, for the obtain of a more holistic picture of the antioxidant landscape, can be followed through the implementation of High-Resolution (HR) techniques, i.e. conducting analysis of anthocyanins and phenolic compounds via UHPLC-Orbitrap-MS [22, 23], or anthocyanins and other phenolic compounds' analysis (phenolic acids, ellagitannins, and flavonoids) in pomegranate juices via HPLC-DAD-ESI-qTOF-MS [24]. Additionally, HRMS targeted and untargeted analysis in conjunction with chemometrics can be used alongside of bioactive compound determination, as well as a reliable tool for pomegranate adulteration [25]. Additionally, pomegranates' metabolite profiling and implementation of chemometrics was conducted via Nuclear Magnetic Resonance (NMR) spectroscopy [26].

The requirement for developing novel workflows capable of handling the massive amount of data derived from HRMS has emerged. Vendor specific and open access software have been utilized to interpret the acquired data; however, the scientific community has noted the significance of employing and evaluating open-source software due to the variety of algorithms, the files' compatibility between vendors, the codes' transparency, the large community of software developers and the capacity for their modification according to various licensing schemes. Nevertheless, there are still issues using non-targeted MS data derived either from Data Independent Acquisition (DIA-bbCid) or Data dependent (DDA-automs). DDA and DIA modes were employed in conjunction, to maximize the benefits of each mode and ensure the mining of the largest features' number. On the one hand, DDA is the most used strategy for compound elucidation, due to its cleaner and more easily interpretable spectra [27]. On the other hand, since DIA detects and fragments all ions in a sample, it empowers more thorough and repeatable analysis by collecting data within a wide range of known and unknown ions, while the fragmentation spectra are more complicated in interpretation [28].

In the terms of this project, innovative antioxidant pomegranate-based juices have been produced. In this direction, this research serves two supplementary purposes. The first aim is the molecular characterization of the drink *per se* in terms of quantitative and qualitative constitution. The second aim is the development of an advanced mass spectrometry novel workflow for the comprehensive characterization of the drink in terms of compounds' identification that bares antioxidant activity. This has been accomplished via the compilation of an extensive suspect list of antioxidants for the bioactivity characterization, and the assembling of the literature-based suspect list for the usual comprehensive characterization. It is noteworthy, that the aforementioned bioactivity-based characterization is a novel approach aiming towards the fast and efficient exploitation of the chemical domain. A new role for the combined suspect lists as a searchable database has been highlighted, towards the automated suspect screening, and increasing the credibility of the identification results. Therefore, the bioactive-based and literature-based characterization of foods and beverages may pave the way for a more comprehensive identification, while the utilization of open-source software provides an alternative yet efficient tool for the scientific community.

2. Results

2.1. Suspect Screening of the juice employing different workflows- Qualitative results

In total, 29 compounds were identified in all investigated juices, employing the suspect screening methodology reaching to different identification confidence levels depending on the available information. The levels of identification based on the criteria set on the scientific work of Schymanski et al. [29]. Specifically, 17 compounds reached at the level of identification 1, 10 compounds were identified at level of identification 2a, 1 compound reached to level 2b, and 1 compound was identified at level 3.

The identified compounds belonged to several categories with potential beneficial effect on human health. Specifically, 8 organic acids (citric acid, malic acid, gallic acid, gentistic acid, chlorogenic acid, pyroglutamic acid, fumaric acid, quinic acid), 4 fatty acids (linoleic acid, oleic acid, palmitic acid, linolenic acid), 3 amino acids (phenylalanine, leucine, norvaline) and 1 organic acid ester (ethyl gallate) were identified. Additionally, 10 flavonoids and the metabolites thereof (ellagic acid, ellagic acid glucoside, quercetin, rutin, apigenin, 2-Phenylethyl beta-Dglucopyranoside, kaempferol, phlorizin, verbacoside), 2 sugars (Fructose, glucosamine) were also identified.

All the compounds identified are tabulated in Table 1, where the compound name, the molecular formula, the experimental and the predicted retention time, the theoretical and experimental m/z value of the precursor ion, and the ionization mode are provided. Additionally, the 5 most intense MS/MS fragments (if they existed) of the sample and their corresponding ones obtained either from the spectral library or the reference standards are presented. The cosine similarity scores of the investigated spectra for all samples compared, as acquired from MS-DIAL, and the corresponding level of identification are also presented. The samples are coded as 80%, 90% and 100% based on the percentage of pomgranate, which is the basic ingredient.

Table 1. Compounds identified through suspect screening in the final product.

Compound Name	Chemical Formula	Exp. t _R (min) (Reference Standard) ^a	Pred. t _R (min)	Application Domain ^b	Exp. m/z ^c	Theor. m/z	ESI Mode	MS/MS Explained Fragments ^d	Reference MS/MS Spectra ^e	Total score			Level of Identification/ Database Reference ⁱ
										80% ^f	90% ^g	100% ^h	
Citric acid	C ₆ H ₈ O ₇	1.2 (1.2)	1.1	Box 1	191.0213	191.0197	-ESI	57.0353	57.0354	0.76	0.75	0.76	1
								87.0092	87.0089				
								111.0094	111.0088				
								191.0198	191.0199				
Malic acid	C ₄ H ₆ O ₅	1.2 (1.1)	1.0	Box 1	133.0131	133.0143	-ESI	71.01442	71.0139	0.69	0.72	0.65	1
								115.00472	115.0022				
								133.01266	133.0138				
Fructose	C ₆ H ₁₂ O ₆	1.4	1.7	Box 1	179.05754	179.0561	-ESI	89.0224	89.0246	0.66	0.64	0.69	1
								179.0560	179.0558				
Gallic acid	C ₇ H ₆ O ₅	1.6 (1.5)	2.9	Box 2	169.0151	169.0142	-ESI	69.0354	69.0346	0.66	0.66	0.76	1
								97.0271	97.0295				
								125.0245	125.0244				
Gentisic acid	C ₇ H ₆ O ₄	2.2 (2.4)	3.0	Box 1	153.0199	153.0193	-ESI	108.0191	108.0217	0.70	0.70	0.76	1
								109.0289	109.0295				
Chlorogenic acid	C ₁₆ H ₁₈ O ₉	2.9 (2.9)	3.5	Box 1	353.0896	353.0878	-ESI	161.0264	161.0233	0.74	0.76	0.76	1
								173.0479	173.0447				
								191.0557	191.0555				
								192.0564	192.0589				
Fumaric Acid	C ₄ H ₄ O ₄	1.3 (1.2)	1.9	Box 1	115.0035	115.0037	-ESI	71.0136	71.0136	0.68	0.71	0.74	2a MzCloud no 1274
								72.9928	72.9925				
								115.0035	115.004				
Quinic acid	C ₇ H ₁₂ O ₆	1.3 (1.3)	1.2	Box 1	191.0561	191.0561	-ESI	85.0291	85.0299	0.62	0.64	0.69	1
								191.0564	191.0558				
Phenylalanine	C ₉ H ₁₁ NO ₂	3.0	4.2	Box 2	164.0726	164.0717	-ESI	72.0091	72.0099	0.63	0.66	0.61	1
								147.0447	147.0448				
								164.0728	164.0712				

Compound Name	Chemical Formula	Exp. t_R (min) (Reference Standard) ^a	Pred. t_R (min)	Application Domain ^b	Exp. m/z ^c	Theor. m/z	ESI Mode	MS/MS Explained Fragments ^d	Reference MS/MS Spectra ^e	Total score			Level of Identification/ Database Reference ⁱ
										80% ^f	90% ^g	100% ^h	
Leucine	C ₆ H ₁₃ NO ₂	2.8	1.8	Box 1	132.1020	132.1019	+ESI	58.0654	58.0634	0.69	0.68	0.69	1
								69.0697	69.0686				
								86.0964	86.0956				
								87.0989	87.0987				
Norvaline	C ₅ H ₁₁ NO ₂	4.5	1.2	Box 3	118.0646	118.0863	+ESI	132.1019	132.1013	0.71	0.73	0.72	2a MonA ID: FiehnHILIC002191
								65.03932	65.036				
								117.0584	117.057				
								118.0658	118.062				
Quercetin	C ₁₅ H ₁₀ O ₇	7.3 (7.3) ^a	7.0	Box 1	301.0361	301.0354	-ESI	151.0042	151.0037	0.68	0.61	0.70	1
								169.0170	169.0135				
								179.0004	178.9996				
Rutin	C ₂₇ H ₃₀ O ₁₆	5.7 (5.7) ^a	6.2	Box 1		609.1461	-ESI	300.028	300.0256	0.62	0.63	0.68	1
								301.0312	301.0366				
								117.0359	117.0341				
Apigenin	C ₁₅ H ₁₀ O ₅	8.3 (7.9) ^a	7.6	Box 1	269.0460	269.0455	-ESI	151.0077	151.0029	0.61	0.66	0.61	1
								269.0463	269.0459				
								133.0305	133.0297				
Kaempferol	C ₁₅ H ₁₀ O ₆	7.6 (7.8) ^a	7.2	Box 1	285.042	285.0405	-ESI	151.0030	151.0039	0.76	0.78	0.71	2a GNPS ID: VF-NPL- QEHF014174
								175.0386	175.0388				
								285.0421	285.0400				
								161.0319	161.0244				
Verbascoside	C ₂₉ H ₃₆ O ₁₅	5.0 (4.8) ^a	8.4	Box 4	623.1993	623.1981	-ESI	162.0263	162.0278	0.62	0.69	0.68	1
Phloridzin	C ₂₁ H ₂₄ O ₁₀	5.9 (5.8) ^a	8.2	Box 2	435.1291	435.1297	-ESI	167.0362	167.0340	0.66	0.67	0.75	1

Compound Name	Chemical Formula	Exp. tr (min) (Reference Standard) ^a	Pred. tr (min)	Application Domain ^b	Exp. m/z ^c	Theor. m/z	ESI Mode	MS/MS Explained Fragments ^d	Reference MS/MS Spectra ^e	Total score			Level of Identification/ Database Reference ⁱ
										80% ^f	90% ^g	100% ^h	
Ethyl gallate	C ₉ H ₁₀ O ₅	4.9 (5.0)	5.0	Box 1	197.0473	197.0455	-ESI	123.0061	123.0086	0.62	0.63	0.62	3 FoodB ID: FDB012004
								140.0102	140.0118				
								168.0066	168.0074				
								169.0149	169.0146				
Linoleic acid	C ₁₈ H ₃₂ O ₂	13.5 (13.5) ^a	12.9	Box 1	279.2336	279.2330	-ESI	279.2327	279.2328	0.61	0.60	0.70	1
								280.2345	280.2333				
Oleic acid	C ₁₈ H ₃₄ O ₂	14.0 (14.0) ^a	13.3	Box 1	281.2486	281.2486	-ESI	281.2484	281.2468	0.69	0.69	0.74	1
								282.2526	282.2508				
Palmitic acid	C ₁₆ H ₃₂ O ₂	13.8 (13.8) ^a	13.0	Box 1	255.2334	255.2330	-ESI	255.2329	255.2327	0.84	0.83	0.86	1
								256.2366	256.2364				
								257.2349	257.2395				
Linolenic acid	C ₁₈ H ₃₀ O ₂	13.0	12.4	Box 1	277.2170	277.2173	-ESI	277.2198	277.2180	0.91	0.92	0.95	2a MoNA ID: MetaboBASE0976
Ellagic acid	C ₁₄ H ₆ O ₈	4.6	4.7	Box 1	300.9993	300.9990	-ESI	201.0183	201.0200	0.77	0.74	0.80	2a MoNA ID: FiehnHILIC001170
								229.0143	229.0144				
								283.9960	283.9950				
								299.9924	299.9900				
Glucosamine	C ₆ H ₁₃ NO ₅	1.8	1.4	Box 1	162.0764	162.0760	+ESI	60.0450	60.0443	0.94	0.97	0.93	2a GNPS ID: CCM-SLIB00005464276
								72.0450	72.0435				
								84.0450	84.0445				
								85.0290	85.0284				
2-Phenylethyl beta-D-glucopyranoside	C ₁₄ H ₂₀ O ₆	6.5	6.2	Box 1	302.1616	302.1600	+ESI	162.0760	162.0744	0.89	0.92	0.94	2a GNPS ID: CCMSLIB00000854907
								81.0337	81.0330				
								85.0287	85.0270				
								97.0289	97.0280				
								105.0707	105.0710				
127.0340	127.0400												

Compound Name	Chemical Formula	Exp. t_R (min) (Reference Standard) ^a	Pred. t_R (min)	Application Domain ^b	Exp. m/z ^c	Theor. m/z	ESI Mode	MS/MS Explained Fragments ^d	Reference MS/MS Spectra ^e	Total score			Level of Identification/ Database Reference ^f
										80% ^f	90% ^g	100% ^h	
Pyroglutamic acid	C ₅ H ₇ NO ₃	2.4	2.1	Box1	130.0511	130.0507	+ESI	84.0455	84.0460	0.94	0.93	0.91	2a MassBank ID: PR311148
								85.0483	85.0450				
								129.0190	129.0220				
								130.0508	130.0510				
4-Hydroxyquinoline	C ₉ H ₇ NO	4.5	6.1	Box 2	146.0606	146.0600	+ESI	77.0389	77.03700	0.93	0.97	0.99	2a RIKEN PLaSMA ID: RIKENPlasma000824
								91.0541	91.0560				
								101.0395	101.0440				
								146.0598	146.0610				
Dihydrozeatin	C ₁₀ H ₁₅ N ₅ O	5.8	4.31	Box 2	222.1351	222.1349	+ESI	69.0699	69.0710	0.91	0.88	0.92	2a MoNA ID: FiehnHILIC000308
								136.0615	136.0620				
								148.0626	148.0620				
								204.1227	204.1250				
								222.1347	222.1349				
Ellagic acid glucoside	C ₂₀ H ₁₆ O ₁₃	3.7	4.5	Box 1	463.0514	463.0518	-ESI	300.9976					2b diagnostic ion Using Smilib

^a retention time of the reference standard, ^b Development and Prediction of Retention Time Indices on-line available at <http://rti.chem.uoa.gr> ^c experimental m/z value with error ± 0.005 Da; $[M + H]^+$ for +ESI and $[M - H]^-$ for -ESI (*with the exception of glucosamine, which m/z value corresponds to $[M - H_2O + H]^+$, and 2-phenylethyl beta-D-glucopyranoside which m/z corresponds to $[M + NH_4]^+$), ^d top-five most intense explained peaks (if they existed), ^e MS/MS fragments of the spectra reference standard or mass spectral library, ^f 80% pomegranate, ^g 90% pomegranate, ^h 100% pomegranate, ⁱ for the level of identification 2a the database entry is referred.

170
171
172
173
174
175

2.2. Quantitative analysis

The compounds, for which their analytical standards were available in the laboratory, were quantified using calibration curves of reference standards. Specifically, the concentrations of 5 organic acids (abscisic acid, chlorogenic acid, citric acid, gallic acid, quinic acid), 2 flavonoids (quercetin, galangin), one flavonoid glucoside (verbascoside), and phenol glucoside (phlorizin) were determined. The compounds catechin, gentistic acid, epicatechin, genistein, p-coumaric, and pinobanksin were identified through target screening; however their concentrations were below their limit of quantification (LOQ) defined as 0.5 mg/kg. The abovementioned concentrations with their corresponding standard deviation (SD) for the three investigated samples are presented. Additionally, the corresponding calibration equations in the form of $y = (a \pm S_a) x + (b \pm S_b)$, as well as their correlation coefficients are tabulated in Table 2.

Table 2. Quantification results.

Analyte	Concentration (mg/kg) \pm SD (n=3)	Concentration (mg/kg) \pm SD (n=3)	Concentration (mg/kg) \pm SD (n=3)	Equation of the external calibration curve $y = (a \pm S_a) x + (b \pm S_b)$	Correlation coefficient R^2
	80% ^a	90% ^b	100% ^c		
Abscisic acid	0.28 \pm 0.02	<LOQ	<LOQ	$y = (29407 \pm 1108) x + (13932 \pm 5655)$	0.994
Chlorogenic acid	4.07 \pm 0.33	1.35 \pm 0.08	0.52 \pm 0.05	$y = (299437 \pm 17948) x + (236799 \pm 91609)$	0.98
Citric acid	203 \pm 18.9	204 \pm 21.2	199 \pm 18.4	$y = (114212 \pm 3871) x - (27802 \pm 19759)$	0.991
Galangin	1.41 \pm 0.86	0.65 \pm 0.05	<LOQ	$y = (179124 \pm 11301) x - (9729 \pm 57678)$	0.98
Gallic acid	5.72 \pm 0.41	4.57 \pm 0.41	5.11 \pm 0.47	$y = (46623 \pm 3535) x + (57556 \pm 18043)$	0.98
Phloridzin	1.01 \pm 0.09	0.65 \pm 0.05	0.65 \pm 0.06	$y = (304319 \pm 25920) x + (287534 \pm 132294)$	0.97
Quinic acid	0.75 \pm 0.09	0.38 \pm 0.04	<LOQ	$y = (121371 \pm 1970) x + (431844 \pm 10055)$	0.993
Verbascoside	0.87 \pm 0.12	0.51 \pm 0.05	<LOQ	$y = (58119 \pm 1053) x - (14120 \pm 5376)$	0.996
Quercetin	13.1 \pm 0.45	11.6 \pm 0.56	11 \pm 0.48	$y = (71193 \pm 4859) x + (51014 \pm 24802)$	0.992

^a 80% pomegranate, ^b 90% pomegranate, ^c 100% pomegranate

3. Discussion

3.1. Development of a novel workflow

The necessity for the development of a novel workflow that combines targeted and suspect screening with the existing DDA and DIA fragmentation methodologies emerged lately. In the current data treatment software landscape, only MS-DIAL has the appropriate algorithms to perform both DDA and DIA (MS2Dec, CorrDec) analysis. On the other hand, MS-DIAL is not designed to perform target screening, whereas MZmine [30] is capable of performing target screening based on MS1 spectra and annotation based only on DDA fragmentation. Therefore, novel workflows that combined these two pieces of software together were designed to overcome these issues.

The DDA and DIA approaches, producing fragmentation of the molecular species, differ essentially to the precursor selection. Thus, in DDA, the precursor ion is selected, followed by fragmentation, whereas in DIA, no precursor ion is selected, instead, all ions are fragmented. Therefore, in a formal way only DDA produces MS/MS spectra, while DIA generates MS/MS like fragmentation but in MS1 spectra. Therefore, for the rest of the

manuscript the term “fragmentation derived spectra” will be used to describe the fragmentation pattern derived from either DDA (MS/MS) or DIA (high collision energy).

3.1.2 Compilation of suspect lists

In the direction of deeply mining all the potential information according to their antioxidant contribution, a novel idea was conceived called bioactivity driven interrogation. Therefore, focusing on a desirable property of the final product, substances of a specified activity were highlighted, bypassing the fuzzy information concerning the whole metabolite landscape. A new list that filtered the specific biological activity of the chemical space, i.e., emphasizing in the current case to the antioxidant capacity (the antioxidant active compounds) has been assembled. The compiled list has been entitled as “Bioactivity Driven Suspect List (BDSL)”.

Taking into consideration the metabolism of the most abundant antioxidant compounds of the BDSL, another list was compiled through **combinatorial enumeration** in order to predict products from potential metabolic pathways, such as the glucolysation and methylation. This assembled list, called Virtual Metabolite Suspect List (**VMSL**), was generated using Smilib v2.0 [31] [32], utilizing the scaffolds, linkers and building blocks according to the software. The scaffolds are the already identified Natural Products (NPs) of the BDSL, while as building blocks have been used one or two glucose units and/or a methyl group. Additionally, to retain the parent compounds, hydrogen has been also selected as a building block. It should be noted that the metabolites of the compounds thereof could potentially have enhanced biological activity compared to their initial non-metabolized counterparts (i.e., quercetin may have similar antioxidant capacity with quercetin glucosides or even enhanced, due to the latter’s different hydrophilicity). Alongside with the aforementioned suspect lists, a literature-based list (LBL) has also been compiled.

3.1.3 Mzmine-based workflow (MS1 driven)

Mzmine has a targeted feature detection module, which can interrogate the MS1 experiment using the information from a suspect list and construct the respective Extracted Ion Chromatograms (EICs), as the feature list. These chromatographic peaks were prioritized based on the area under curve. It has to be reminded that the antioxidants need to be in high quantities in order to exert their role. Thus, the most abundant antioxidants (MAA) were selected and used as input to construct a searchable database. This list will be utilized as a part of the MS-DIAL workflow. Therefore, these two lists also function in a confirmatory way as the simultaneous presence of a metabolite enhances the confidence. Thus, 6 common compounds were identified from both lists (VMSL and LBL), i.e., quercetin, kaempferol, apigenin, gentisic acid, gallic acid and chlorogenic acid.

3.1.4 MS-DIAL based workflow (fragmentation driven)

MS-DIAL deems to be a valuable solution, in order to exploit the DIA results, as well as DDA spectra. The compounds are annotated offline by comparing the DDA or DIA deconvoluted spectra to the corresponding ones from the samples. MS-DIAL searches against already assembled local libraries (i.e., the general list ESI(+/-)-MS/MS assembled from authentic standards, which is provided from the software’s download page). A novel idea has been conceived about the replacement of the abovementioned library from a narrowed version, encompassing only specific compounds of interest (i.e., target/suspect version). In contrast to the untargeted mode, for which MS-DIAL has originally been used, this novel approach allows the software to function in the target/suspect mode. Therefore, this flexibility allows the construction of custom-made libraries, providing the capacity to narrow down the number of plausible candidates.

MSP files (editable with a simple text editor i.e., notepad, work pad etc.) consist of entries that include the candidates' names, the molecular formula, the exact mass, the theoretical retention time, and the MS/MS fragments. Such files are publicly available from various sources such as the GNPS, MS-DIAL etc. webpages. These MSP files were adjusted to focus on the analytes of interest, thus serving as a database, which is essentially a suspect list. This approach offers the additional advantage of a more complete view for spectra comparison. Thus, the fragments included in this database correspond to experimental spectra and not biased/curated fragments as it happens commonly for the compilation of suspect lists. These files are compatible with the MS-DIAL software, which offers the potential for process both DDA and DIA data. Two lists have been generated (LBL, VM SL) and imported to MS-DIAL. The overall chemical space was searched with the aid of these two suspect lists, aiming to find the bioactive content in terms of antioxidants, and secondly to chemically characterize in a comprehensive way the final drinks.

3.2. Comparative analysis of antioxidant juices

As the main aim of this endeavor was the development of a drink with enhanced antioxidant activity, various combinations of raw materials in different percentages (i.e., 3.3% ,6.6% from other juices) have been used. For clarification purposes, it should be reminded that the secondary ingredients used; were juices of aronia, black berry and red grapes. The contribution of the quantity for the selected antioxidants has been studied. Generally, 3 patterns have been observed based either to the targeted results or the corresponding peak areas (used for the substances, which their reference standards were not available). An increasing trend of the investigated antioxidants when the percentage of other juices was lowered (i.e., quercetin), an opposite effect (i.e., ethyl gallate), or no effect (i.e., fructose). Thus, in the case of ethyl gallate, the drinks containing higher percentage of the secondary ingredients, also contain higher amounts of this substance comparing to the pure pomegranate drink. On the other hand, the amount of quercetin in juices (containing 80 and 90% pomegranate) is lower. Finally, the same amount of fructose has been determined in all three analyzed juices. This is depicted in the figure 1. No discrepancies for these patterns were noticed, which validates the results of the analysis.

The compounds quinic acid, kaempferol, quercetin, chlorogenic acid, rutin, verbacoscide were found higher in the juice supplemented with 6.66 % of each secondary raw material. Their elevated quantity is connected to the enhanced antioxidant activity.

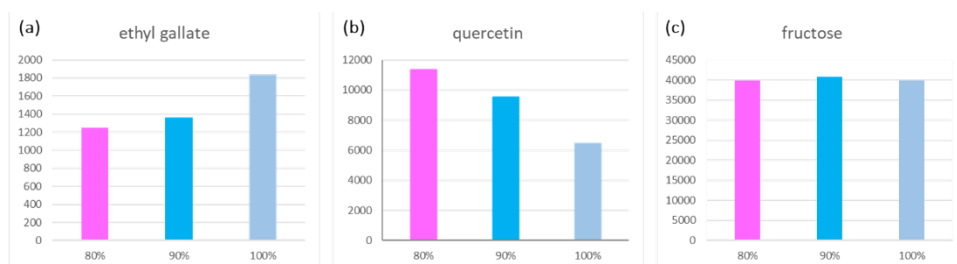


Figure 1: Quantity of (a) ethyl gallate, (b) quercetin (c) fructose for the designed juices with 80%, 90%, 100% of pomegranate.

3.3. Beneficial role of the identified compound in humans' health

One prevalent criterion for the selection and the final percent contribution of each ingredient to the produced drink is their antioxidant capacity in terms of the existence and content of bioactive substances. Hence, the presence of the antioxidants should be adequately high, as their activity is not excreted by the interaction of the substance with a pharmacological target/receptor. The obtained results revealed the presence of the antioxidants compounds belonging to several categories (organic acids, fatty acids, amino acids,

flavonoids and metabolites, etc.). The beneficial effect of the most important identified compounds is briefly discussed.

Ellagic acid is a well-known antioxidant that has been shown to be effective in preventing neurodegeneration by repairing mitochondrial damage and scavenging free radicals [33]. Quercetin is a powerful antioxidant known for its capacity to prevent tissue damage [34]. Kaempferol has anticarcinogenic, antioxidant and anti-inflammatory [35], as well as antibacterial, antifungal, and antiprotozoal activities [36].

Fumaric acid has anti-inflammatory, neuroprotective, chemo preventive activities [37], and acts against multiple sclerosis (MS) [38]. Chlorogenic acid has antioxidant antibacterial, hepatoprotective, cardioprotective, anti-inflammatory, antipyretic, neuroprotective, anti-obesity, antiviral, anti-microbial, anti-hypertension activity [39]. Citric acid is a secondary antioxidant [40], yielding synergistically to enhance primary antioxidants' activity [41], while its chelating and acidulating properties (25) are well-known [42].

4. Materials and Methods

4.1. Methodology of preparation Pomegranate based drink

A thorough literature review was conducted for 16 potential raw materials that might serve as additional ingredients in the pomegranate-based juice to enhance its nutritional value. The raw materials investigated were *Prunus cerasifera*, *Vaccinium vitis-idaea* L, *Prunus cerasus*, *Aronia melanocarpa*, *Citrus*, *Ribes rubrum*, *Vitis vinifera* L, *Hippophae*, *Actinidia deliciosa*, *Opuntia ficus-Indica*, *Ficus carica*, *Rubus occidentalis*, *Morus alba*, *Morus nigra*.

The *Morus nigra* (black berries), *Aronia melanocarpa* (aronia), and *Vitis vinifera* L (red grapes) were chosen based on their market availability, antioxidant contribution, and effect on the flavor and the lingering of the final product.

Then, two mixtures of juices were created, only differing in the proportion of their ingredients. Specifically, the % percentage of pomegranate, black berries, aronia, red grapes were (90, 3.33, 3.33, 3.33, v/v) and (80, 6.66, 6.66, 6.66, v/v), respectively. Their flavor (sweetness, sour taste, acidity), lingering flavor (sour taste, acidity) as well as color, fragrance, and texture have been assessed. The second mentioned juice had a higher overall score in the majority of the investigated categories.

The manufacturing process of the final product was initialized with the defrosting of the raw components (juices of pomegranate, red grape, black berry and aronia) until they reached at room temperature. Afterwards, the transfer of the juices to tanks and their combination through stirring followed. Next, a pasteurization step in a tube heat exchanger at 83 °C was performed. Next, hot filling took place, and the bottles were sealed using an automatic sealing machine. The juices' temperature has been decreased in a cooling tunnel. The bottles were kept at a temperature of 20 °C, protected from light.

4.2. Reagents and Materials

All standards and reagents used were of analytical grade (<95%), unless differently stated explicitly. Methanol (MeOH, LC-MS grade) was purchased from Merck (Darmstadt, Germany), while formic acid 99% and acetic acid were acquired from Fluka (Buchs, Switzerland). Ammonium acetate and ammonium formate were obtained from Fisher Scientific (Geel, Belgium). The ultrapure water (H₂O) was provided by a Milli-Q device (Millipore Direct-Q UV, Bedford, MA, USA). Regenerated cellulose syringe filters (RC filters, pore size 0.2 µm, diameter 15 mm) were acquired from Macherey-Nagel (Düren, Germany). Citric acid, chlorogenic acid, gallic acid, malic acid, quercetin, apigenin, phloridzin, ethyl gallate, L-phenylalanine, leucine, fatty acid methylesters, D (-) fructose, catechin, epicatechin, pinobanksin and p-coumaric acid were obtained from Sigma Aldrich (Stenheim, Germany). Verbascoside was purchased from HWI pharma services (Rülzheim, Germany), while quinic acid, genistein, gentistic acid, and galangin were acquired from

Supelco (Stenheim, Germany). Apigenin was purchased from Alfa Aesar (Karlsruhe, Germany). 350
351

Stock solutions of the reference standards (1000 mg L⁻¹) were prepared in MeOH (LC- 352
MS grade) and stored at -20 °C in amber glass vials. A solution of 50 mg L⁻¹ was prepared 353
by appropriate dilution of the individual stock standard solutions. Following that, dilu- 354
tions with a mixture of MeOH: H₂O (80:20, v/v) were carried out in order to prepare work- 355
ing solutions with concentrations of 0.5, 1, 2.5, 5, 10 mg L⁻¹. 356
357

4.3. Sample Pre-Treatment for HRMS Analysis 358

In an eppendorf tube, 200 mg of the drink were weighed and the addition of 200 µL 359
MeOH: H₂O (80:20, v/v) was followed. The mixture was vortexed vigorously and filtered 360
through RC syringe filters. The extracts were transferred to 2-mL autosampler glass vials 361
and injected into the UPLC-QToF-MS system in both ionization modes. 362
363

4.4. Instrumentation 364

4.4.1. UPLC-QToF-MS Instrumentation 365

The chemical analysis of the pomegranate-based juice was carried out using Ultra- 366
High-Pressure Liquid Chromatography-Quadruple Time of Flight Mass Spectrometry 367
(UPLC-QToF-MS) employed with an HPG-3400 pump (Dionex Ultimate 3000 RSLC, 368
Thermo Fisher Scientific, Dreieich, Germany) coupled to a time-of-flight mass analyzer 369
(Hybrid Quadrupole time of Flight Matic Bruker Daltonics, Bremen, Germany). The chro- 370
matographic column utilized was an Acclaim RSLC 120 C18 column (2.2 µm, 2.1 × 100 371
mm, Thermo Fisher Scientific, Dreieich, Germany), equipped with a pre-column (Van 372
guard Acquity UPLC BEH C18 (1.7 µm, 2.1 × 5 mm, Waters, Ireland) and its temperature 373
(30 °C) was maintained constant during the analysis. In the positive ionization mode, the 374
mobile phases consisted of (A) aq. 5 mM ammonium formate: MeOH (90:10, v/v) acidified 375
with 0.01% formic acid and (B) 5 mM ammonium formate in MeOH acidified with 0.01% 376
formic acid. In the negative ionization mode, the mobile phases were (a) aq. 10 mM am- 377
monium acetate: MeOH (90:10, v/v) and (B) 10 mM ammonium acetate in MeOH. The 378
same gradient elution program was used in both ionization modes. The gradient program 379
is described in detail in a previous work of our group [43]. The values selected for the MS 380
parameters were capillary voltage of 3500 V, nebulizer gas pressure of 2 bar (N₂), drying 381
gas flow rate of 8 L min⁻¹, and capillary temperature of 200 °C. The sodium formate cali- 382
brant, which was prepared in H₂O: isopropanol (50:50, v/v), was injected at the beginning 383
of each run to calibrate the Q-ToF system on a daily basis. 384

4.5. Mass Spectrometry Data Analysis 385

4.5.1. Identification Confidence 386

The feature annotation was performed according to the Schymanski et al. scheme, 387
considering the five levels of confidence in identifying a plausible candidate [29]. At level 388
5, the only confirmed information is the exact mass of interest, while there is no infor- 389
mation concerning its molecular mass. At level of identification 4, the candidate's molec- 390
ular formula is confirmed [44], whereas at the next level (level 3), the tentatively identi- 391
fication via evaluation of candidates' MS/MS fragment is realized utilizing *in silico* fragmen- 392
tation tools (MetFrag [45] or CFM-ID [46]). Additionally, at this identification level, prior- 393
itization methods, such as retention time prediction [47] and ionization efficiency estima- 394
tion [48] can be used to enhance the identification confidence. In the cases, in which diag- 395
nostic ion exist, the plausible candidate can reach at level identification 2b. At level 396
identification 2a, potential candidates can reach, when the corresponding MS/MS spectra are 397
available at spectral libraries and their similarity score is higher than 0.7. At level of 398
identification 1, the candidates' reference standards are available and their MS/MS spectra, the 399
retention time being in accordance. 400

4.5.2 Data processing and identification workflows

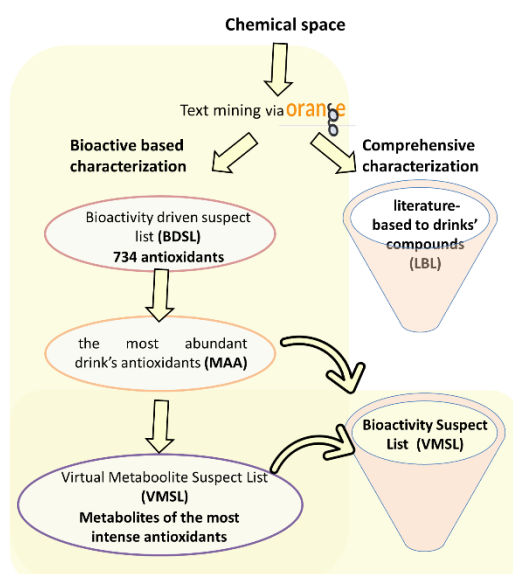
4.5.2.1 Workflows for the compilation of suspect lists.

4.5.2.1.1 Bioactivity driven suspect list.

A suspect list compiled of 734 antioxidant substances was retrieved using Orange statistical language (version 3.33.0) through the text mining module using PubChem data. The molecular formula and the exact mass, alongside with the compound name, were deposited in a csv file, which in turn was uploaded to MZmine 2.53. The raw data were calibrated and converted to mzxml files using Data Analysis software (Bruker Daltonics, Bremen, Germany) to be compatible with MZmine. The most abundant substances i.e., those with the highest chromatographic peak areas were selected for the evaluation of the drink's antioxidant capacity. The mass spectral databases used for the assembling of the suspect list were: MoNa [49], MassBank-Europe [50], METLIN, Human Metabolome Database (HMDB) [51], and the Global Natural Products Social Molecular Networking (GNPS) [52]. The features were annotated through the comparison of their MS/MS spectra with the corresponding ones from the spectral libraries or the reference standards in the cases that they were available in the laboratory. Due to the lack of entries concerning the metabolites derived from the enumeration process as well as their MS/MS spectra from the aforementioned libraries, their fragmentation was estimated based solely on the characteristic diagnostic ions (i.e., for ellagic glucoside, the fragment of the aglucone part and the corresponding fragment of the sugar moiety). These pieces of information were also added to the VMSL list.

4.5.2.1.2. Comprehensive literature based suspect list.

An exhaustive literature-based, text-mining-defined suspect list was created using the Orange statistical language. This list encompasses the compounds retrieved from PubChem that were specified for pomegranate, black berries, red grapes, and aronia. In the same direction, a literature-based suspect list [2, 20-26, 53-55] was assembled with the traditional way and merged with the one obtained from the text mining procedure. This list has been used for the suspect/target screening protocol and as a supporting tool to enhance the confidence of the acquired results from the BDSL. Furthermore, these two workflows act synergistically to provide a holistic picture of the plant's chemical composition. The workflows employed for the compilation of the suspect lists is illustrated to figure 2.



401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

432

433

434

435

436

437

Figure 2. Workflow of suspect lists compilation.

4.5.2.2 Methodology of the development of Workflow MS1 Driven

The raw data were calibrated, converted to mzxml files and uploaded to MZmine 2.53. The list of 734 antioxidants in csv form were imported and the ones with the highest peak area derived from the feature list were used as scaffolds for the compilation of VMSL.

4.5.2.3 Methodology of the development of Workflow MS2 Driven

The calibrated raw data were converted to abf files (ABF converter) [56], and then uploaded to the open-source MS-DIAL software (version 4.92) [57]. Both acquisition modes, DDA and DIA, were examined. DDA was selected for the most abundant compounds, whereas DIA was utilized for the compounds found in lower quantities. The different acquisition modes have been processed separately.

Based on the compounds mentioned in the literature (LBL), an in-house database was created and imported into MS-DIAL (MSP file format). Additionally, the VMSL (MSP file format) was also imported to MS-DIAL. These two MSP files were processed separately to evaluate the antioxidant content, as well as the compounds discovered through the comprehensive drinks' characterization. The online "Retention time prediction tool" (available on <http://rti.chem.uoa.gr/>) was utilized to predict the theoretical retention time of each compound by uploading the canonical smiles. For compounds with reference standards not available in the laboratory, the corresponding spectra were retrieved from public spectral libraries. A procedure blank was also prepared. The chromatographic peak areas of the procedure blank have to be 5-fold lower than the ones in the sample, in order not to be excluded as false positives.

The entire workflow is depicted in figure 3.

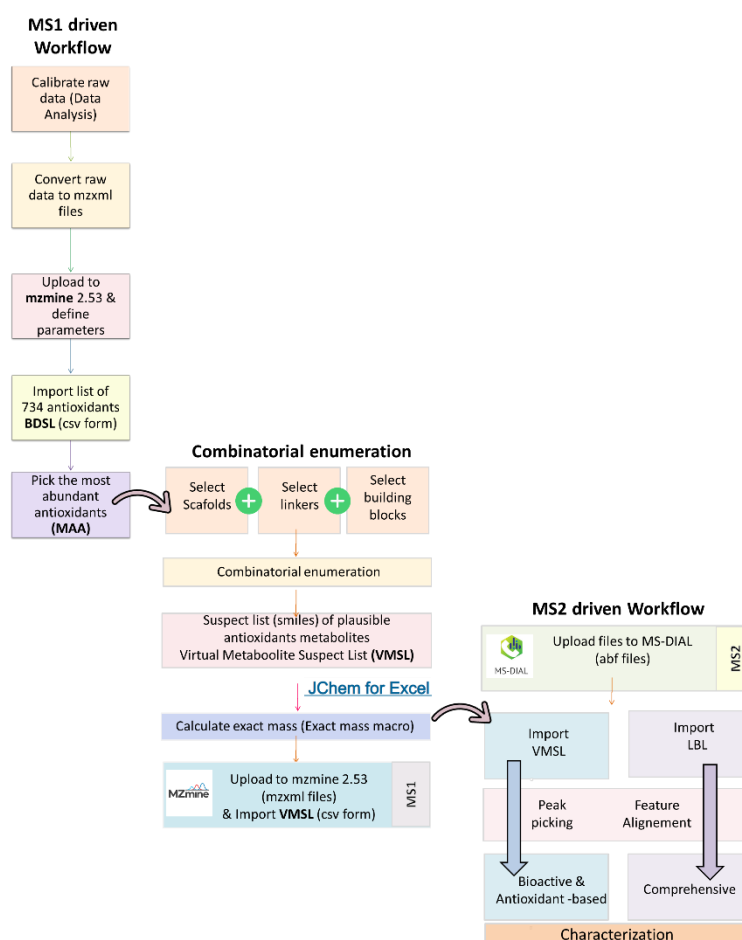


Figure 3. Workflow of the novel suspect screening methodology

4.6. Target screening Methodology

For the determination of the compound's concentration, TASQ 1.4 (Bruker Daltonics, Bremen, Germany) was used. For the compounds with available reference standards, and belonging in the category of bioactive compounds, their quantification was followed. The quantification of the analytes was performed using standard calibration curves and based on a validated method developed in our laboratory [25]. Satisfactory linearity was achieved for all the analytes (R^2 values ranging from 0.97 to 0.996).

5. Conclusions

Pomegranate-based juices with antioxidant capacity has been designed, produced, and characterized employing novel suspect and target screening methodologies through open-source software using UPLC-QToF-MS. A total of 29 compounds, including fatty acids, amino acids, organic acids, flavonoids and their metabolites were identified in the drinks via the developed methodologies in both ionization modes. The significant amount of quercetin, as well as the high concentration of citric acid sparked a lot of interest, due to their plausible positive impact on human health.

In this context, novel suspect, and target screening methodologies for the elucidation of drinks' compounds have been developed to ascertain a faster and less effort-consuming data treatment, ensuring results with enhanced credibility. Bioactivity-/combinatorial- and literature-based lists have been assembled as searchable database in combination to the mass spectrometry analysis using open-source software (MZmine, MS-DIAL).

Author Contributions: Conceptualization, N.S.T.; methodology, A.P., E.G; validation, A.P., E.G.; data curation: A.P; investigation: A.P., I.T; formal analysis, A.P., I.T; resources, N.S.T.; writing—original draft preparation, A.P.; E.G, writing—review and editing, A.P., N.S.T., and E.G.; supervision; N.S.T. and E.G.; project administration, N.S.T.; funding acquisition, N.S.T. All authors have read and agreed to the published version of the manuscript.

Funding: This research was co-financed by the European Regional Development Fund of the European Union and Greek national funds through the call Research and Innovation Strategies for Smart Specialisation (RIS3) (project code: AMOP7-0072324).

Institutional Review Board Statement: Not applicable

Informed Consent Statement: Not applicable

Data Availability Statement: Data sharing not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds are not available from the authors.

References

1. Sreekumar, S.; Sithul, H.; Muraleedharan, P.; Azeez, J. M.; Sreeharshan, S., Pomegranate fruit as a rich source of biologically active compounds. *Biomed Res Int* **2014**, 2014, 686921 DOI: 10.1155/2014/686921.
2. Maria I. Gil, F. A. T.-B., Betty Hess-Pierce, Deirdre M. Holcroft, and Adel A. Kader, Antioxidant Activity of Pomegranate Juice and Its Relationship with Phenolic Composition and Processing. *J. Agric. Food Chem.* **2000**, 48, 4581–4589.
3. Viuda-Martos, M.; El Gendy Ael, N.; Sendra, E.; Fernandez-Lopez, J.; Abd El Razik, K. A.; Omer, E. A.; Perez-Alvarez, J. A., Chemical composition and antioxidant and anti-Listeria activities of essential oils obtained from some Egyptian plants. *Journal of agricultural and food chemistry* **2010**, 58, (16), 9063-70 DOI: 10.1021/jf101620c.
4. Elfalleh, W., Total phenolic contents and antioxidant activities of pomegranate peel, seed, leaf and flower. *Journal of Medicinal Plants Research* **2012**, 6, (32), DOI: 10.5897/jmpr11.995.
5. Gurčík, L.; Bajusová, Z.; Ladvenicová, J.; Palkovič, J.; Novotná, K., Cultivation and Processing of Modern Superfood—Aronia melanocarpa (Black Chokeberry) in Slovak Republic. *Agriculture* **2023**, 13, (3), DOI: 10.3390/agriculture13030604.
6. Gajic, D.; Saksida, T.; Koprivica, I.; Vujicic, M.; Despotovic, S.; Savikin, K.; Jankovic, T.; Stojanovic, I., Chokeberry (Aronia melanocarpa) fruit extract modulates immune response in vivo and in vitro. *Journal of Functional Foods* **2020**, 66, DOI: 10.1016/j.jff.2020.103836.
7. Hussain, F.; Rana, Z.; Shafique, H.; Malik, A.; Hussain, Z., Phytopharmacological potential of different species of Morus alba and their bioactive phytochemicals: A review. *Asian Pacific Journal of Tropical Biomedicine* **2017**, 7, (10), 950-956 DOI: 10.1016/j.apjtb.2017.09.015.
8. Ercisli, S.; Orhan, E., Chemical composition of white (Morus alba), red (Morus rubra) and black (Morus nigra) mulberry fruits. *Food Chemistry* **2007**, 103, (4), 1380-1384 DOI: 10.1016/j.foodchem.2006.10.054.
9. Kamiloglu, S.; Serali, O.; Unal, N.; Capanoglu, E., Antioxidant activity and polyphenol composition of black mulberry (Morus nigra L.) products. *Journal of Berry Research* **2013**, 3, (1), 41-51 DOI: 10.3233/jbr-130045.
10. Integrated Taxonomic Information System (ITIS). Vol. 2023.
11. Bigard, A.; Berhe, D. T.; Maoddi, E.; Sire, Y.; Boursiquot, J. M.; Ojeda, H.; Peros, J. P.; Doligez, A.; Romieu, C.; Torregrosa, L., Vitis vinifera L. Fruit Diversity to Breed Varieties Anticipating Climate Changes. *Frontiers in plant science* **2018**, 9, 455 DOI: 10.3389/fpls.2018.00455.
12. Wolkovich, E. M.; García de Cortázar-Atauri, I.; Morales-Castilla, I.; Nicholas, K. A.; Lacombe, T., From Pinot to Xinomavro in the world's future wine-growing regions. *Nature Climate Change* **2018**, 8, (1), 29-37 DOI: 10.1038/s41558-017-0016-6.
13. Sabra, A.; Netticadan, T.; Wijekoon, C., Grape bioactive molecules, and the potential health benefits in reducing the risk of heart diseases. *Food Chem X* **2021**, 12, 100149 DOI: 10.1016/j.fochx.2021.100149.

14. Pomegranate Market Share and Forecast till 2028. <https://www.marketwatch.com/press-release/pomegranate-market-share-and-forecast-till-2028-2023-03-19> (22/03/2023), 537
538
15. DEJIAN HUANG, B. O., AND RONALD L. PRIOR, The Chemistry behind Antioxidant Capacity Assays. *J. Agric. Food Chem.*, 53, **2005**, 53, 1841–1856. 539
540
16. Sendra, J. M.; Sentandreu, E.; Navarro, J. L., Reduction kinetics of the free stable radical 2,2-diphenyl-1-picrylhydrazyl (DPPH•) for determination of the antiradical activity of citrus juices. *European Food Research and Technology* **2006**, 223, (5), 615–624 DOI: 10.1007/s00217-005-0243-3. 541
542
543
17. de la Fuente, B.; Pallares, N.; Barba, F. J.; Berrada, H., An Integrated Approach for the Valorization of Sea Bass (*Dicentrarchus labrax*) Side Streams: Evaluation of Contaminants and Development of Antioxidant Protein Extracts by Pressurized Liquid Extraction. *Foods* **2021**, 10, (3), DOI: 10.3390/foods10030546. 544
545
546
18. NAVINDRA P. SEERAM, M. A., YANJUN ZHANG, SUSANNE M. HENNING, LYDIA FENG, MARK DREHER; HEBER, A. D., Comparison of Antioxidant Potency of Commonly Consumed Polyphenol-Rich Beverages in the United States. *J. Agric. Food Chem.* **2008**, 56, 1415–1422. 547
548
549
19. Gozlekci, S.; Saracoglu, O.; Onursal, E.; Ozgen, M., Total phenolic distribution of juice, peel, and seed extracts of four pomegranate cultivars. *Pharmacogn Mag* **2011**, 7, (26), 161–4 DOI: 10.4103/0973-1296.80681. 550
551
20. Ehling, S.; Cole, S., Analysis of organic acids in fruit juices by liquid chromatography-mass spectrometry: an enhanced tool for authenticity testing. *Journal of agricultural and food chemistry* **2011**, 59, (6), 2229–34 DOI: 10.1021/jf104527e. 552
553
554
21. Akhavan, H.; Barzegar, M.; Weidlich, H.; Zimmermann, B. F., Phenolic Compounds and Antioxidant Activity of Juices from Ten Iranian Pomegranate Cultivars Depend on Extraction. *Journal of Chemistry* **2015**, 2015, 1–7 DOI: 10.1155/2015/907101. 555
556
557
22. Di Stefano, V.; Scandurra, S.; Pagliaro, A.; Di Martino, V.; Melilli, M. G., Effect of Sunlight Exposure on Anthocyanin and Non-Anthocyanin Phenolic Levels in Pomegranate Juices by High Resolution Mass Spectrometry Approach. *Foods* **2020**, 9, (9), DOI: 10.3390/foods9091161. 558
559
560
23. Indelicato, S.; Houmanat, K.; Bongiorno, D.; Ejjilani, A.; Hssaini, L.; Razouk, R.; Charafi, J.; Ennahli, S.; Hanine, H., Freeze dried pomegranate juices of Moroccan fruits: main representative phenolic compounds. *J Sci Food Agric* **2023**, 103, (3), 1355–1365 DOI: 10.1002/jsfa.12229. 561
562
563
24. Gomez-Caravaca, A. M.; Verardo, V.; Toselli, M.; Segura-Carretero, A.; Fernandez-Gutierrez, A.; Caboni, M. F., Determination of the major phenolic compounds in pomegranate juices by HPLC-DAD-ESI-MS. *Journal of agricultural and food chemistry* **2013**, 61, (22), 5328–37 DOI: 10.1021/jf400684n. 564
565
566
25. Dasenaki, M. E.; Drakopoulou, S. K.; Aalizadeh, R.; Thomaidis, N. S., Targeted and Untargeted Metabolomics as an Enhanced Tool for the Detection of Pomegranate Juice Adulteration. *Foods* **2019**, 8, (6), DOI: 10.3390/foods8060212. 567
568
569
26. Hasanpour, M.; Saberi, S.; Iranshahi, M., Metabolic Profiling and Untargeted ¹H-NMR-Based Metabolomics Study of Different Iranian Pomegranate (*Punica granatum*) Ecotypes. *Planta medica* **2020**, 86, (3), 212–219 DOI: 10.1055/a-1038-6592. 570
571
572
27. Zhang, C.; Zuo, T.; Wang, X.; Wang, H.; Hu, Y.; Li, Z.; Li, W.; Jia, L.; Qian, Y.; Yang, W.; Yu, H., Integration of Data-Dependent Acquisition (DDA) and Data-Independent High-Definition MS(E) (HDMS(E)) for the Comprehensive Profiling and Characterization of Multicomponents from *Panax japonicus* by UHPLC/IM-QTOF-MS. *Molecules* **2019**, 24, (15), DOI: 10.3390/molecules24152708. 573
574
575
576
28. Géhin, C.; Holman, S. W., Advances in high-resolution mass spectrometry applied to pharmaceuticals in 2020: A whole new age of information. *Analytical Science Advances* **2021**, 2, (3–4), 142–156 DOI: 10.1002/ansa.202000149. 577
578
29. Schymanski, E. L.; Jeon, J.; Gulde, R.; Fenner, K.; Ruff, M.; Singer, H. P.; Hollender, J., Identifying small molecules via high resolution mass spectrometry: communicating confidence. *Environmental science & technology* **2014**, 48, (4), 2097–8 DOI: 10.1021/es5002105. 579
580
581
30. Schmid, R.; Heuckeroth, S.; Korf, A.; Smirnov, A.; Myers, O.; Dyrland, T. S.; Bushuiev, R.; Murray, K. J.; Hoffmann, N.; Lu, M.; Sarvepalli, A.; Zhang, Z.; Fleischauer, M.; Dührkop, K.; Wesner, M.; Hoogstra, S. J.; Rudt, E.; Mokshyna, O.; Brungs, C.; Ponomarov, K.; Mutabdzija, L.; Damiani, T.; Pudney, C. J.; Earll, M.; Helmer, P. O.; Fallon, T. R.; Schulze, T.; Rivas-Ubach, A.; Bilbao, A.; Richter, H.; Nothias, L. F.; Wang, M.; Oresic, M.; Weng, J. K.; Bocker, S.; Jeibmann, A.; Hayen, H.; Karst, U.; Dorrestein, P. C.; Petras, D.; Du, X.; Pluskal, T., Integrative 582
583
584
585
586

- analysis of multimodal mass spectrometry data in MZmine 3. *Nat Biotechnol* **2023**, DOI: 10.1038/s41587-023-01690-2. 587
588
31. SMIlib v2.0. <http://melolab.org/smilib/> (13/04/2023), 589
32. Schüller, A.; Hähnke, V.; Schneider, G., SMIlib v2.0: A Java-Based Tool for Rapid Combinatorial Library Enumeration. *QSAR & Combinatorial Science* **2007**, *26*, (3), 407-410 DOI: 10.1002/qsar.200630101. 590
591
33. Ahmed Touqeer, S. N. W., Nabavi Fazel Seyed, Orhan Erdogan Ilkay, Braidy Nady, Sobarzo-Sanchez Eduardo and Nabavi Mohammad Seyed, Insights Into Effects of Ellagic Acid on the Nervous System: A Mini Review, . 592
<https://dx.doi.org/10.2174/1381612822666160125114503>. **2016**, *22*, (10), 1350-1360 DOI: 593
10.2174/1381612822666160125114503. 594
595
34. Anand David, A. V.; Arulmoli, R.; Parasuraman, S., Overviews of Biological Importance of Quercetin: A Bioactive Flavonoid. *Pharmacogn Rev* **2016**, *10*, (20), 84-89 DOI: 10.4103/0973-7847.194044. 596
597
35. Wang, J.; Fang, X.; Ge, L.; Cao, F.; Zhao, L.; Wang, Z.; Xiao, W., Antitumor, antioxidant and anti-inflammatory activities of kaempferol and its corresponding glycosides and the enzymatic preparation of kaempferol. *PLoS one* **2018**, *13*, (5), e0197563 DOI: 10.1371/journal.pone.0197563. 598
599
600
36. Periferakis, A.; Periferakis, K.; Badarau, I. A.; Petran, E. M.; Popa, D. C.; Caruntu, A.; Costache, R. S.; Scheau, C.; Caruntu, C.; Costache, D. O., Kaempferol: Antimicrobial Properties, Sources, Clinical, and Traditional Applications. *International journal of molecular sciences* **2022**, *23*, (23), DOI: 10.3390/ijms232315054. 601
602
603
37. Reis, F. S.; Stojković, D.; Soković, M.; Glamočlija, J.; Ćirić, A.; Barros, L.; Ferreira, I. C. F. R., Chemical characterization of *Agaricus bohusii*, antioxidant potential and antifungal preserving properties when incorporated in cream cheese. *Food Research International* **2012**, *48*, (2), 620-626 DOI: 604
10.1016/j.foodres.2012.06.013. 605
606
607
38. Bjelobaba Ivana, S. D. a. L. I., Multiple Sclerosis and Neuroinflammation: The Overview of Current and Prospective Therapies. *Current Pharmaceutical Design* **2017**, *23*, 693-730 DOI: 608
10.2174/1381612822666161214153108. 609
610
39. Naveed, M.; Hejazi, V.; Abbas, M.; Kamboh, A. A.; Khan, G. J.; Shumzaid, M.; Ahmad, F.; Babazadeh, D.; FangFang, X.; Modarresi-Ghazani, F.; WenHua, L.; XiaoHui, Z., Chlorogenic acid (CGA): A pharmacological review and call for further research. *Biomedicine & Pharmacotherapy* **2018**, *97*, 67-74 DOI: 611
10.1016/j.biopha.2017.10.064. 612
613
614
40. Ryan, E. M.; Duryee, M. J.; Hollins, A.; Dover, S. K.; Pirruccello, S.; Sayles, H.; Real, K. D.; Hunter, C. D.; Thiele, G. M.; Mikuls, T. R., Antioxidant properties of citric acid interfere with the uricase-based measurement of circulating uric acid. *Journal of pharmaceutical and biomedical analysis* **2019**, *164*, 460-466 DOI: 615
10.1016/j.jpba.2018.11.011. 616
617
618
41. Kilel, E. C.; Wanyoko, J. K.; Faraj, A. K.; Ngoda, P., Effect of Citric Acid on the Total Monomeric Anthocyanins and Antioxidant Activity of Liquor Made from Unprocessed Purple Leafed TRFK 306 Kenyan Tea Clone. *Food and Nutrition Sciences* **2019**, *10*, (10), 1191-1201 DOI: 10.4236/fns.2019.1010086. 619
620
621
42. H. ROSTAMZAD, B. S., M. KASHANINEJAD and A. SHABANI, ANTIOXIDATIVE ACTIVITY OF CITRIC AND ASCORBIC ACIDS AND THEIR PREVENTIVE EFFECT ON LIPID OXIDATION IN FROZEN PERSIAN STURGEON FILLETS. **2011**, *41*, 135-140. 622
623
624
43. Panara, A.; Gikas, E.; Thomaidis, N. S., From By-Products to Fertilizer: Chemical Characterization Using UPLC-QToF-MS via Suspect and Non-Target Screening Strategies. *Molecules* **2022**, *27*, (11), DOI: 625
10.3390/molecules27113498. 626
627
44. Kind, T.; Fiehn, O., Seven Golden Rules for heuristic filtering of molecular formulas obtained by accurate mass spectrometry. *BMC bioinformatics* **2007**, *8*, 105 DOI: 10.1186/1471-2105-8-105. 628
629
45. Ruttkies, C.; Schymanski, E. L.; Wolf, S.; Hollender, J.; Neumann, S., MetFrag relaunched: incorporating strategies beyond in silico fragmentation. *Journal of cheminformatics* **2016**, *8*, 3 DOI: 10.1186/s13321-016-0115-9. 630
631
46. Djombou-Feunang, Y.; Pon, A.; Karu, N.; Zheng, J.; Li, C.; Arndt, D.; Gautam, M.; Allen, F.; Wishart, D. S., CFM-ID 3.0: Significantly Improved ESI-MS/MS Prediction and Compound Identification. *Metabolites* **2019**, *9*, (4), DOI: 10.3390/metabo9040072. 632
633
634
47. Aalizadeh, R.; Nika, M.-C.; Thomaidis, N. S., Development and application of retention time prediction models in the suspect and non-target screening of emerging contaminants. *Journal of Hazardous Materials* **2019**, *363*, 277-285 DOI: <https://doi.org/10.1016/j.jhazmat.2018.09.047>. 635
636
637

48. Krauss, M.; Singer, H.; Hollender, J., LC-high resolution MS in environmental analysis: from target screening to the identification of unknowns. *Analytical and bioanalytical chemistry* **2010**, *397*, (3), 943-51 DOI: 10.1007/s00216-010-3608-9. 638
639
49. MoNA, M. o. N. A. <http://mona.fiehnlab.ucdavis.edu/> 642
50. MassBank-Europe <https://massbank.eu/MassBank/> 643
51. Wishart, D. S.; Guo, A.; Oler, E.; Wang, F.; Anjum, A.; Peters, H.; Dizon, R.; Sayeeda, Z.; Tian, S.; Lee, B. L.; Berjanskii, M.; Mah, R.; Yamamoto, M.; Jovel, J.; Torres-Calzada, C.; Hiebert-Giesbrecht, M.; Lui, V. W.; Varshavi, D.; Varshavi, D.; Allen, D.; Arndt, D.; Khetarpal, N.; Sivakumaran, A.; Harford, K.; Sanford, S.; Yee, K.; Cao, X.; Budinski, Z.; Liigand, J.; Zhang, L.; Zheng, J.; Mandal, R.; Karu, N.; Dambrova, M.; Schioth, H. B.; Greiner, R.; Gautam, V., HMDB 5.0: the Human Metabolome Database for 2022. *Nucleic Acids Res* **2022**, *50*, (D1), D622-D631 DOI: 10.1093/nar/gkab1062. 644
645
646
647
648
52. GNPS <https://gnps.ucsd.edu/ProteoSAFe/libraries.jsp> 649
53. Poyrazoğlu, E.; Gökmen, V.; Artık, N., Organic Acids and Phenolic Compounds in Pomegranates (*Punica granatum* L.) Grown in Turkey. *Journal of Food Composition and Analysis* **2002**, *15*, (5), 567-575 DOI: 10.1006/jfca.2002.1071. 650
651
652
54. Maria I. Gil, J. C., Naceur Ayed, Francisco Artes, Francisco A. Tomfis-Barberan, Influence of cultivar, maturity stage and geographical location on the juice pigmentation of Tunisian pomegranates. *Z Lebensm Unters Forsch* **1995**, *201*, 361-364. 653
654
655
55. Labbé, M.; Ulloa, P. A.; López, F.; Sáenz, C.; Peña, Á.; Salazar, F. N., Characterization of chemical compositions and bioactive compounds in juices from pomegranates (Wonderful, Chaca and Codpa) at different maturity stages. *Chilean journal of agricultural research* **2016**, *76*, (4), 479-486 DOI: 10.4067/s0718-58392016000400012. 656
657
658
56. Reifycs Abf Converter. <https://www.reifycs.com/AbfConverter> 659
57. Tsugawa, H.; Cajka, T.; Kind, T.; Ma, Y.; Higgins, B.; Ikeda, K.; Kanazawa, M.; VanderGheynst, J.; Fiehn, O.; Arita, M., MS-DIAL: data-independent MS/MS deconvolution for comprehensive metabolome analysis. *Nature methods* **2015**, *12*, (6), 523-6 DOI: 10.1038/nmeth.3393. 660
661
662
663